

<https://doi.org/10.59854/dhrrh.2024.2.2.103>

– CASE REPORT –

Cutaneous Complications after Autologous Stem Cell Transplantation

Ana-Maria ILINESCU¹, Ana-Maria VLADAREANU¹, Elena Georgiana ENE¹,
Iuliana IORDAN^{1,2}, Horia BUMBEA^{1*}

Abstract

Autologous stem cell transplantation (ASCT) has become more prevalent for high-grade non-Hodgkin's lymphoma (NHL) over the past decade. ASCT serves as a salvage treatment for high-risk or relapsed patients, offering potential long-term remission. Cutaneous modifications post-ASCT can be classified as early (within 100 days) and late (after 100 days). Understanding these changes is crucial for timely diagnosis and management. Early modifications typically include transient erythema, hyperpigmentation, and acute dermatitis, often linked to the conditioning regimen. Late modifications, occurring months to years post-transplantation, can encompass chronic skin changes such as scleroderma-like features and persistent pigmentary alterations. Emerging evidence challenges the notion that ASCT is free of graft-versus-host disease (GVHD), with incidences of auto-GVHD reported. However, cutaneous GVHD post-ASCT, though uncommon, poses significant diagnostic challenges. Cutaneous GVHD manifestations range from mild eruptions to severe, systemic involvement. The role of skin biopsy in these cases is invaluable, providing histopathological confirmation and distinguishing GVHD from other dermatoses. Recognizing the early and late cutaneous modifications following ASCT is essential for clinicians. Despite the rarity of skin GVHD in ASCT, awareness and prompt biopsy can facilitate accurate diagnosis and appropriate management, improving patient outcomes.

¹Hematology Department, Carol Davila University of Medicine and Pharmacy; Emergency University Hospital, Bucharest, Romania

²Medical Semiology Department, Carol Davila University of Medicine and Pharmacy, Emergency University Hospital, Bucharest, Romania
epartment of Hematology, Emergency Municipal Hospital Timisoara, Romania

Corresponding author:

* **Professor Horia Bumbea**, Hematology Department, Carol Davila University of Medicine and Pharmacy; Emergency University Hospital, Bucharest, Romania
email: horiabum@gmail.com

Introduction

Autologous hematopoietic stem cell transplantation (ASCT), developed in the early 1980s, represents a major advancement in treating aggressive lymphoproliferative disorders. The procedure involves administering chemotherapy to induce remission and mobilize autologous stem cells into the peripheral blood, followed by stem cell collection. Patients then receive high-dose chemotherapy to eradicate residual malignant cells, and

finally, the autologous stem cells are infused to "rescue" or restore hematopoiesis.

The use of ASCT to treat high grade non-Hodgkin's lymphoma (NHL) has increased over the past decade. While intensive chemotherapy can achieve disease free survival (DFS) rates of up to 50% at five years in certain subtypes of aggressive lymphoma, the likelihood of a favorable response is significantly reduced in high-risk patients and those with relapsed or refractory disease.

ASCT can serve as a salvage treatment option in these scenarios. Retrospective analyses suggest that it can lead to long-term progression-free survival and even cure in a proportion of patients. This evidence, combined with increased availability and experience with ASCT, has led to changes in standard medical practice. More hematologists are now referring patients for transplant, and patient eligibility criteria are gradually broadening.

The Paradigm study conducted by the Blood and Marrow Transplant Clinical Trials Network (BMT CTN) and the Cancer and Leukemia Group B (CALGB) is currently the largest randomized study comparing ASCT with chemoimmunotherapy in any NHL subtype. Preliminary results indicate that ASCT consolidation is particularly beneficial for individuals with high-risk or relapsed disease. Therefore, with this study as well as other recent studies showing that transplantation in first remission is superior to that performed at a later time, ASCT looks to become increasingly utilized in the treatment of diffuse large B cell lymphoma (DLBCL).

The ASCT procedure carries significant risks and is generally reserved for fit, relatively young patients. Although transplant-related mortality has decreased in recent years, it remains at approximately 5-10% for patients undergoing ASCT for lymphoma. A substantial portion of this risk is due to infection, disease relapse or progression, secondary primary malignancies, organ failure, and treatment-related toxicity.

Cutaneous complications in ASCT

Cutaneous complications following ASCT can vary from mild to life-threatening, with a wide range of skin issues

reported. These complications are categorized into early and late, depending on their onset relative to the transplantation. Early complications occur within the first 100 days post-transplant, while late complications arise after 100 days. The underlying mechanisms differ between these two categories, which will be discussed in further detail.

Clinical scenario

A 40-year-old patient was diagnosed with stage IIIA DLBCL not otherwise specified (NOS), germinal center B cell-like (GCB) subtype. The patient was initially treated with R-CHOP immunochemotherapy. Upon disease relapse, he received radiotherapy and R-DHAP, followed by ASCT in partial remission. The pre-transplant conditioning regimen included Lomustine, Etoposide, Cytarabine, and Melphalan. The bone graft, consisting of 37.06×10^6 CD34+ cells, was administered. Grafting occurred on the 12th day. The post-transplant period was complicated by persistent fever of unknown origin and vasovagal syncope.

On day +27 post-transplantation, the patient presented to the Emergency Department with altered general status, loss of appetite, nausea, persistent high fever, and generalized pruritic maculopapular exanthema (Figure 1). Laboratory investigations showed elevated liver enzymes and a positive cytomegalovirus (CMV) PCR (viral load 172.01 copies/ml). Broad-spectrum antibiotics, Valganciclovir, and corticosteroid pulse-therapy were initiated. The patient's clinical condition promptly improved, with resolution of fever and disappearance of the rash.

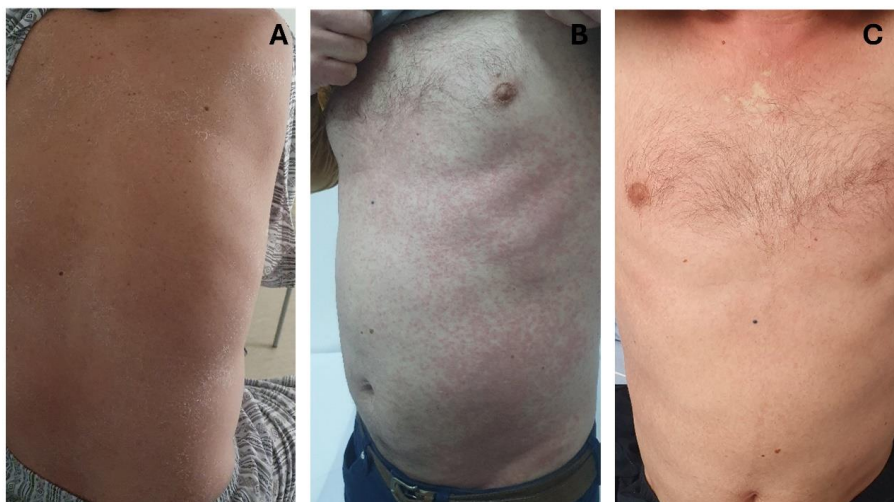


Figure 1. Evolution of the generalised maculo-papular exanthema following ASCT at three time points: A. Day +27; B. Day +49; C. Day +62

Early cutaneous complications following ASCT

The timeline for symptoms onset is further divided into two periods: the pre-engraftment period (the first 30 days) and the post-engraftment period (30 days or more). During the pre-engraftment period, high-dose chemotherapy causes mucosal damage and immunosuppression, leading to numerous infectious cutaneous complications. In the post-engraftment period, characterized by immune system reconstitution, the patients can suffer from acute graft-versus-host disease (GVHD) and treatment-related cutaneous complications.

The differential diagnosis is often challenging and typically requires a skin biopsy for accurate identification. Our patient developed early cutaneous complications following ASCT, occurring around the borderline of the two time periods (before/after day +30). The primary challenge was establishing a differential diagnosis. We considered acute GVHD, cutaneous complications due to CMV reactivation, and engraftment syndrome as potential causes.

Differential diagnosis

The pathologies that should be considered in the differential diagnosis of cutaneous complications are presented in Table 1.

Pathology	Clinical presentation	Histological findings
<i>Acute GVHD</i>	<i>Grade 0-4 Head, neck, acral sites (described below in detail)</i>	<i>(described below in detail)</i>
<i>Drug reaction</i>	<i>Morbilliform erythema Onset 1-2 or 7-10 days after exposure to new drug Begins on torso and spreads the next days</i>	<i>May not involve hair follicles. Fewer eosinophils</i>
<i>Viral infections (HHV6, HHV7, CMV)</i>	<i>Morbilliform or maculopapular eruptions</i>	<i>Rare perivascular lymphocytic infiltrates Intranuclear inclusions – CMV</i>
<i>Acral erythema</i>	<i>Secondary to chemotherapy Affects extremities (palms, soles) Edema, blistering, desquamation, pain</i>	<i>Dermatitis, necrotic and dyskeratotic keratinocytes, edema of dermis, lymphocytes infiltrate in perivascular space, with vacuolar degeneration</i>
<i>Toxic epidermal necrolysis</i>	<i>Involves more than 30% body surface area Appears after 1-3 weeks after exposure to new drug Prodrome of flu-like symptoms Skin eruption: macular rash, bullae, blisters; Nikolsky sign Affect face and torso</i>	<i>Necrosis and epidermis detachment Few mononuclear cells Satellitosis High eosinophilic necrosis</i>
<i>Engraftment syndrome</i>	<i>Self-limited skin exanthem Transient non-infectious fever, diarrhea, pulmonary infiltration/ edema, impaired renal/ liver function, transient encephalopathy, weight gain</i>	<i>Eosinophilic necrosis of epidermal keratinocytes in epidermis Vacuolization in the basal layer Lymphocytic infiltration of perivascular space</i>
<i>Radiation dermatitis</i>	<i>Erythema, desquamation, blisters, necrosis</i>	<i>Dermal sclerosis, elastosis, vascular ectasia, hyperkeratosis, epidermal spongiosis, vacuolization of basal layer; hyperchromasia of stroma fibroblasts and endothelial cells, enlargement and atypia, inflammatory infiltrate</i>
<i>Lupus erythematosus</i>	<i>Malar rash on cheeks and nose Rash of the forehead, neck with sparing of nasolabial folds Edema, papules Lasting days-weeks, may appear after sun exposure Telangiectasis, erosion, dyspigmentation, poikiloderma</i>	<i>Fibrinoid necrosis, liquefactive degenerations and atrophy of the epidermis, edema, hemorrhages, lymphocytic infiltrate in dermis</i>

Table 1. Differential diagnosis between acute GVHD and other skin conditions after ASCT
Abbreviations: GVHD=graft-versus-host disease, HHV=human herpesvirus, CMV=Cytomegalovirus

Graft-versus-host disease

While GVHD is traditionally associated with allogeneic stem cell transplantation, its occurrence after ASCT is becoming more recognized. GVHD involves an immune-mediated process where donor T lymphocytes attack recipient tissues, leading to a multisystem disorder. In allogeneic transplantation, GVHD has been associated with a graft-versus-lymphoma effect, contributing to improved disease control. However, GVHD also correlates with increased morbidity and mortality. Studies involving mixed patient population have indicated poorer survival outcomes in those experiencing GVHD complication.

Although ASCT has generally been considered to be free of GVHD due to the absence of donor immune cells, emerging evidence suggests otherwise. In the ASCT, the GVHD is a form of “auto-aggression syndrome” (auto-GVHD). Auto-GVHD has been reported to arise unpredictably or after immunomodulation therapy. A growing number of case reports have highlighted the occurrence of a chronic form of GVHD following ASCT, even in the absence of donor cells.

A comprehensive study conducted by Copelan et al. sheds light on the incidence rate of GVHD following ASCT in DLBCL patients. This study included 402 DLBCL patients. The overall GVHD incidence rate was 39%, with 27% experiencing acute GVHD and 20% chronic GVHD. These results align with findings from other smaller studies. A retrospective analysis by the Center for International Blood and Marrow Transplant Research reported a 36% GVHD incidence rate, while another single-center study reported a rate of 28%.

The clinical manifestation of cutaneous GVHD ranges from erythematous maculopapular morbilliform eruption and follicular erythema, to erythroderma and epidermolysis.⁴⁰ It may impact areas like the face, ears, palms, soles, extending to the trunk, or even the entire body surface area. Patients may be asymptomatic or report itching or dysesthesia. Additionally, atypical presentations such as pityriasis rubra pilaris, acquired ichthyosis, and psoriasis vulgaris-like eruptions are also possible.

The most common changes encountered in cutaneous GVHD are:

- Poikiloderma: atrophy, hypopigmentation, hyperpigmentation as patches, telangiectasias
- Lichen planus-like features: erythematous or violaceous papules/ plaques on dorsal hands and feet, forearms, trunk; scaling is possible

- Sclerosis: wavy aspect and thickening of fibrous septa in fat, on medial arms and thighs; fascial affliction – contractures determining impaired function of limb
- Morphea-like features: plaques of various colors; shiny appearance of skin; hair loss
- Lichen sclerosis-like features

Depending on the clinical features of the rash and its spread, skin GVHD can be classified into the following grades:

0. Without active rash

1. Maculopapular rash involving less than 25% body surface area
2. Maculopapular rash involving 25-50% body surface area
3. Maculopapular rash involving more than 25% body surface area
4. Generalized erythema involving >50% body surface area, bullae/ desquamation involving more than 5% body surface area

The positive diagnosis relies on several factors, including the affected sites (such as the face, palms, and soles), dermoscopic changes (pink or red background and telangiectasias). Additionally, the emergence of rashes following sun exposure is a distinct indicator of skin GVHD. Performing a skin biopsy is essential for the positive diagnosis.

The histological changes found in acute cutaneous GVHD are heterogeneous. The most common findings, according to the National Institute of Health Consensus, are:

ducts, and/or lichenoid inflammation, vacuolar change, or lymphocytic satellitosis;

- Lichen planus-like features: epidermal orthohyperparakeratosis, hypergranulosis, acanthosis, lichenoid inflammation, and/or vacuolar changes of eccrine units;
- Lichen sclerosus-like features: uniformity and/or sclerosis of collagen in papillary dermis, melanophages in papillary dermis, rare lymphocytic infiltration;
- Morphea-like features: thickening and homogenization of collagen clumps in reticular dermis and/or pandermal sclerosis;
- Fasciitis: thickened fascial septa, inflammation, and/or subcutis sclerosis.⁵¹

In our scenario, to confirm the diagnosis, we conducted a skin biopsy. Due to logistical constraints, the biopsy was performed five days after initiating corticosteroid therapy. Histopathological analysis indicated nonspecific changes, including orthokeratosis with areas of parakeratosis,

occasional necrotic keratinocytes, thickened dermis, and perivascular inflammatory infiltrates. Immunohistochemical assessment demonstrated negativity for CD20 and CD79a. These nonspecific changes could potentially be interpreted as GVHD-related, considering the high-dose steroids administered before the biopsy. Furthermore, the changes resolved with steroids treatment.

Evolution and management

The management approach for GVHD is dictated by its classification, grading, extent of involvement, and symptoms. The treatment options vary, ranging from oral antihistamines and moisturizers, to short-time topical steroids and topical calcineurin inhibitors in grade I GVHD. For grades II-IV, methylprednisolone and calcineurin inhibitors are commonly used. Second-line therapies may include extracorporeal photopheresis, mycophenolate mofetil, TNF-antagonists (such as infliximab and etanercept) particularly for cases with gastrointestinal involvement, IL2 receptor monoclonal antibodies (daclizumab), and anti-thymocyte globulins.

In our specific case, despite initially responding well to corticosteroid treatment, the patient returned to the Emergency Department on day +49 with deteriorating clinical condition and high fever. Laboratory tests indicated elevated liver enzymes, although this time the PCR for CMV was negative. Corticosteroid therapy and broad-spectrum antibiotics were restarted. However, the patient experienced complications stemming from corticosteroid treatment, including corticosteroid-induced

diabetes mellitus and depressive disorder with suicidal thoughts. No organic causes for the psychiatric symptoms were identified. Managing these complications involved insulin therapy, antidepressants, antipsychotics, and anxiolytics. Corticosteroid therapy continued for one month. During this period, the rash and itching subsided, and the patient's overall health notably improved.

Conclusion

Diagnosing cutaneous complications following ASCT is challenging due to similar clinical manifestations from various causes. However, since treatment options often vary, establishing an accurate diagnosis is preferable. Skin biopsy is the most valuable tool for this purpose; however, in some cases, including ours, the findings may be nonspecific. Understanding the various causes of skin changes can help guide physicians towards appropriate treatment, even in the absence of definitive data.

No funding for this study

Conflicts of interest

I undersign, certificate that I do not have any financial or personal relationships that might bias the content of this work. The authors declare no conflict of interest.

The authors declare that all the procedures and experiments of this study respect the ethical standards in the Helsinki Declaration of 1975, as revised in 2008(5), as well as the national law. Informed consent was obtained from the patient included in the study.

References

1. Herrera AF, Ahn KW, Litovich C, Chen Y, Assal A, Bashir Q, et al. Autologous and allogeneic hematopoietic cell transplantation for diffuse large B-cell lymphoma-type Richter syndrome. *Blood advances*. 2021;5(18):3528-3539. doi: 10.1182/bloodadvances.2021004865. PMID: 34496026.
2. Ma J, Sun S, Hu Y, Wu M, Shen L, Fulati W, et al. Novel conditioning regimen in upfront autologous stem cell transplantation in high-risk DLBCL. *Bone Marrow Transplantation*. 2022;57(10), 1612-1614. doi: 10.1038/s41409-022-01766-8.
3. Wudhikarn K, Johnson BM, Inwards DJ, Porrata LF, Micallef IN, Ansell SM, et al. Outcomes of Older Adults with Non-Hodgkin Lymphoma Undergoing Autologous Stem Cell Transplantation: A Mayo Clinic

Cohort Analysis. *Transplantation and Cellular Therapy*. 2023;29(3), 176-e1. doi: 10.1016/j.jtct.2022.12.011.

4. Kumar, S., Sharma, A., Pramanik, R., Pathak, N., Gogia, A., Kumar, A., ... & Raina, V. (2022). Long-term outcomes and safety trends of autologous stem-cell transplantation in non-hodgkin lymphoma: a report from a tertiary care center in India. *JCO Global Oncology*, 8(1), e2100383.

5. Wang J, Duan X, Yang L, Liu X, Hao C, Dong H, et al. Comparison of survival between autologous and allogeneic stem cell transplantation in patients with relapsed or refractory B-cell non-Hodgkin lymphoma: a meta-analysis. *Cell Transplantation*. 2020;29, 0963689720975397. doi: 10.1177/0963689720975397.

6. Bentolila G, Pavlovsky A. Relapse or refractory Hodgkin lymphoma: determining risk of relapse or progression after autologous stem-cell transplantation.

- Leukemia & Lymphoma. 2020; 61(7):1548-1554. doi: 10.1080/10428194.2020.1732959.
7. Sureda A, André M, Borchmann P, da Silva MG, Gisselbrecht C, Vassilakopoulos TP, et al. Improving outcomes after autologous transplantation in relapsed/refractory Hodgkin lymphoma: a European expert perspective. *BMC cancer*. 2020; 20(11):1088. doi: 10.1186/s12885-020-07561-2.
 8. Bazarbachi AH, Al Hamed R, Malard F, Bazarbachi A, Harousseau JL, Mohty M. Induction therapy prior to autologous stem cell transplantation (ASCT) in newly diagnosed multiple myeloma: an update. *Blood Cancer Journal*. 2022;12(3):47. doi: 10.1038/s41408-022-00645-1.
 9. Akay OM, Ozbalak M, Pehlivan M, Yildiz B, Uzay A, Yigenoglu TN, Ferhanoglu B. Brentuximab vedotin consolidation therapy after autologous stem-cell transplantation in patients with high-risk Hodgkin lymphoma: Multicenter retrospective study. *Hematological Oncology*, 39(4), 498-505. doi: 10.1002/hon.2897.
 10. Pichereau C, Lengliné E, Valade S, Michonneau D, Ghrenassia E, Lemiale V, Azoulay E. Trajectories of acute graft-versus-host disease and mortality in critically ill allogeneic-hematopoietic stem cell recipients: The Allo-GRRR-OH score. *Bone marrow transplantation*. 2020;55(10), 1966-1974. doi: 10.1038/s41409-020-0857-x
 11. Waszczuk-Gajda A, Penack O, Sbianchi G, Koster L, Blaise D, Reményi P, et al. Complications of Autologous Stem Cell Transplantation in Multiple Myeloma: Results from the CALM Study. *J Clin Med*. 2022;11(12):3541. doi: 10.3390/jcm11123541.
 12. Strunz PP, Froehlich M, Gernert M, Schwaneck EC, Fleischer A, Pecher AC, et al. Immunological Adverse Events After Autologous Hematopoietic Stem Cell Transplantation in Systemic Sclerosis Patients. *Front Immunol*. 2021;12:723349. doi: 10.3389/fimmu.2021.723349.
 13. Kuba A, Raida L. Graft versus Host Disease: From Basic Pathogenic Principles to DNA Damage Response and Cellular Senescence. *Mediators Inflamm*. 2018;9451950. doi: 10.1155/2018/9451950.
 14. Svensson CK, Cowen EW, Gaspari AA. Cutaneous drug reactions. *Pharmacol Rev*. 2001;53(3):357. PMID: 11546834.
 15. Alsaad, K O. My approach to superficial inflammatory dermatoses. *Journal of Clinical Pathology*. 2005;58(12):1233-1241. doi: 10.1136/jcp.2005.027151.
 16. Mays SR, Kunishige JH, Truong E, Kontoyiannis DP, Hymes SR. Approach to the morbilliform eruption in the hematopoietic transplant patient. *Semin Cutan Med Surg*. 2007;26(3):155. doi: 10.1016/j.sder.2007.09.004.
 17. Chidharla A, Kanderi T, Kasi A. Chemotherapy Acral Erythema. [Updated 2023 Feb 27]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK459375/>.
 18. Labib A, Milroy C. Toxic Epidermal Necrolysis. [Updated 2023 May 8]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK574530/>.
 19. Maqbool S, Nadeem M, Shahroz A, Naimat K, Khan I, Tahir H, et al. Engraftment syndrome following Hematopoietic stem cell transplantation: a systematic approach toward diagnosis and management. *Med Oncol*. 2022;40(1):36. doi: 10.1007/s12032-022-01894-7.
 20. Young-Ho L, Yeon JL, Jung-Yum K, Young-Dae K, Seung-Won L. Pre-engraftment syndrome in hematopoietic stem cell transplantation. *Journal of Korean medical science* 2008;23(1):98-103. doi:10.3346/jkms.2008.23.1.98.
 21. Radiation dermatitis pathology | DermNet (dermnetnz.org)
 22. Cooper EE, Pisano CE, Shapiro SC. Cutaneous Manifestations of "Lupus": Systemic Lupus Erythematosus and Beyond. *Int J Rheumatol*. 2021; 2021:6610509. doi: 10.1155/2021/6610509.
 23. Mowafak H. "Skin inflammatory (nontumor) > Lichenoid and interface reaction patterns > Lupus: systemic lupus erythematosus (SLE)". *PathologyOutlines*. Topic Completed: 1 August 2011. Revised: 26 March 2019
 24. Buxbaum, NP, Pavletic SZ. Autoimmunity following allogeneic hematopoietic stem cell transplantation. *Frontiers in Immunology*. 2020;11:2017. doi: 10.3389/fimmu.2020.02017.
 25. Burt RK, Muraro PA, Farge D, Oliveira MC, Snowden JA, Saccardi R, Burman J. New autoimmune diseases after autologous hematopoietic stem cell transplantation for multiple sclerosis. *Bone Marrow*

- Transplantation. 2020;56(7):1509-1517. doi: 10.1038/s41409-021-01277-y.
26. Kumari R, Palaniyandi S, Hildebrandt GC. Metabolic reprogramming—a new era how to prevent and treat graft versus host disease after allogeneic hematopoietic stem cell transplantation has begun. *Frontiers in Pharmacology*. 2020; 11:588449. doi: 10.3389/fphar.2020.588449.
27. Kline J, Subbiah S, Lazarus HM, van Besien K. Autologous graft-versus-host disease: harnessing anti-tumor immunity through impaired self-tolerance. *Bone Marrow Transplant*. 2008;41:505–513. doi: 10.1038/sj.bmt.1705931.
28. Fidler C, Klumpp T, Mangan K, Martin M, Sharma M, Emmons R, Lu M, Kropf P. Spontaneous graft versus host disease occurring in a patient with multiple myeloma after autologous stem cell transplant. *Am J Hematol*. 2012;87:219–221. doi: 10.1002/ajh.22227.
29. Drobyski WR, Hari P, Keever-Taylor C, Komorowski R, Grossman W. Severe autologous GVHD after hematopoietic progenitor cell transplantation for multiple myeloma. *Bone Marrow Transplant*. 2009;43:169–177. doi: 10.1038/bmt.2008.295.
30. Goddard DS, Ruben BS, Mathes ED, Nixon M, Wolf J, Fox LP. A case of severe cutaneous, GI and liver GVHD in a patient with multiple myeloma, status-post-second auto-SCT. *Bone Marrow Transplant*. 2010;45:409–411. doi: 10.4110/in.2013.13.3.107.
31. Krishna SG, Barlogie B, Lamps LW, Krishna K, Aduli F, Anaissie E. Recurrent spontaneous gastrointestinal graft-versus-host disease in autologous hematopoietic stem cell transplantation. *Clin Lymphoma Myeloma Leuk*. 2010;10:17–21. doi: 10.4110/in.2013.13.3.107.
32. Baron F, Gothot A, Salmon JP, Hermanne JP, Pierard GE, Fillet G, Beguin Y. Clinical course and predictive factors for cyclosporin-induced autologous graft-versus-host disease after autologous haematopoietic stem cell transplantation. *Br J Haematol*. 2000;111:745–753. doi: 10.4110/in.2013.13.3.107.
33. Jones RJ, Vogelsang GB, Hess AD, Mann RB, Geller RB, Piantadosi S, Santos GW. Induction of graft-versus-host disease after autologous bone marrow transplantation. *Lancet*. 1989;1:754–757. doi: 10.4110/in.2013.13.3.107.
34. Włodarczyk M, Wachowiak A, Wiczorek K, Toborek M, Wiczorkiewicz-Kabut A, Kata D, Helbig G. Graft-versus-host disease as an unusual complication following autologous stem cell transplantation. *Acta Haematologica Polonica*. 2020;51(1), pp.47-50. DOI:10.2478/ahp-2020-0010.
35. Fidler C, Klumpp T, Mangan K, Martin M, Sharma M, Emmons R, Lu M, Kropf P. Spontaneous graft versus host disease occurring in a patient with multiple myeloma after autologous stem cell transplant. *Am J Hematol*. 2012;87:219–221. doi: 10.1002/ajh.22227.
36. Drobyski WR, Hari P, Keever-Taylor C, Komorowski R, Grossman W. Severe autologous GVHD after hematopoietic progenitor cell transplantation for multiple myeloma. *Bone Marrow Transplant*. 2009;43:169–177. doi: 10.1038/bmt.2008.295.
37. Dietrich S, Dreger P, Hermine O, Kyriakou C, Montoto S, Robinson S, Tanase A. Haploidentical stem cell transplantation for patients with lymphoma: a position statement from the Lymphoma Working Party-European Society for Blood and Marrow Transplantation. *Bone Marrow Transplantation*. 2020;55(2), 317-324.
38. Rey-Búa B, Cabrero M, Bento L, Montoro J, Bastos-Oreiro M, Parody R, García-Sancho AM. Allogeneic Hematopoietic Stem Cell Transplantation in Transformed Follicular Lymphoma (tFL): Results of a Retrospective Multicenter Study from GELTAMO/GETH-TC Spanish Groups. *Cancers*. 2022;14(22), 5670. doi: 10.3390/cancers14225670.
39. Hamadani M, Ngoya M, Sureda A, Bashir Q, Litovich CA, Finel H, Dreger P. Outcome of allogeneic transplantation for mature T-cell lymphomas: impact of donor source and disease characteristics. *Blood Advances*. 2022;6(3),920-930. doi: 10.1182/bloodadvances.2021005899.
40. Filipovich AH, Weisdorf D, Pavletic S, Socie G, Wingard JR, Lee SJ, et al. National Institutes of Health consensus development project on criteria for clinical trials in chronic graft-versus-host disease: I. Diagnosis and staging working group report. *Biol Blood Marrow Transplant*. 2005;11(12):945–956. doi: 10.1016/j.bbmt.2005.09.004.
41. Friedman KJ, LeBoit PE, Farmer ER. Acute follicular graft-vs-host reaction. A distinct clinicopathologic presentation. *Arch Dermatol*. 1988; 124(5):688–691. doi: 10.1001/archderm.1988.01670050032014.
42. Strong Rodrigues K, Oliveira-Ribeiro C, de Abreu Fiuza Gomes S, Knobler R. Cutaneous Graft-Versus-Host

- Disease: Diagnosis and Treatment. *Am J Clin Dermatol.* 2018;(1):33-50. doi: 10.1007/s40257-017-0306-9.
43. Surjana D, Robertson I, Kennedy G, James D, Weedon D. Acute cutaneous graft-versus-host disease resembling type II (atypical adult) pityriasis rubra pilaris. *Australas J Dermatol.* 2015;56(1):e21-e23. doi: 10.1111/ajd.12108.
44. Huang J, Pol-Rodriguez M, Silvers D, Garzon MC. Acquired ichthyosis as a manifestation of acute cutaneous graft-versus-host disease. *Pediatr Dermatol.* 2007;24(1):49-52. doi: 10.1111/j.1525-1470.2007.00333.x.
45. Matsushita T, Hasegawa M, Shirasaki F, Fujimoto M, Yamazaki H, Sato S, et al. A case of acute cutaneous graft-versus-host disease mimicking psoriasis vulgaris. *Dermatology.* 2008;216(1):64-67. doi: 10.1159/000109361.
46. Malard F, Holler E, Sandmaier BM, Huang H, Mohty M. Acute graft-versus-host disease. *Nat Rev Dis Primers.* 2023 Jun 8;9(1):27. doi: 10.1038/s41572-023-00438-1.
47. Hu SW, Cotliar J. Acute graft-versus-host disease following hematopoietic stem-cell transplantation. *Dermatol Ther.* 2011;24(4):411-423. doi: 10.1111/j.1529-8019.2011.01436.x.
48. Kaminska-Winciorek G, Czerw T, Kruzal T, Giebel S. Dermoscopic Follow-Up of the Skin towards Acute Graft-versus-Host-Disease in Patients after Allogeneic Hematopoietic Stem Cell Transplantation. *Biomed Res Int.* 2016;2016:4535717. doi: 10.1155/2016/4535717.
49. Vassallo C, Brazzelli V, Zecca M, Locatelli F, Alessandrino PE, Borroni G. Isomorphic cutaneous graft-versus-host disease reaction after ultraviolet exposure: clinical, histological and direct immunofluorescence studies of four allo-transplanted patients. *J Eur Acad Dermatol Venereol.* 2009;23(8):913-918. doi: 10.1111/j.1468-3083.2009.03220.x.
50. Hillen U, Häusermann P, Massi D, Janin A, Wolff D, Lawitschka A, et al. Consensus on performing skin biopsies, laboratory workup, evaluation of tissue samples and reporting of the results in patients with suspected cutaneous graft-versus-host disease. *J Eur Acad Dermatol Venereol.* 2015;29(5):948-954. doi: 10.1111/jdv.12737.
51. Shulman HM, Cardona DM, Greenson JK, Hingorani S, Horn T, Huber E, et al. Consensus development project on criteria for clinical trials in chronic graft-versus-host disease: II. The 2014 Pathology Working Group Report. *Biol Blood Marrow Transplant.* 2015;21(4):589-603. doi: 10.1016/j.bbmt.2014.12.031.
52. Hymes SR, Alousi AM, Cowen EW. Graft-versus-host disease: part II. Management of cutaneous graft-versus-host disease. *J Am Acad Dermatol.* 2012;66(4):535.e1-535.e16. doi: 10.1016/j.jaad.2011.11.961.
53. Dignan FL, Clark A, Amrolia P, Cornish J, Jackson G, Mahendra P, et al. Diagnosis and management of acute graft-versus-host disease. *Br J Haematol.* 2012;158(1):30-45. doi: 10.1111/j.13652141.2012.09129.x.
54. Jang S, Kim IS, Youn SW. Chronic graft-versus-host disease mimicking psoriasis in a patient with hemophagocytic lymphohistiocytosis. *Ann Dermatol.* 2016;28(1):90-93. doi: 10.5021/ad.2016.28.1.90.
55. Kunitomi A, Lida H, Kamiya Y, Hayashi M, Sao H. Successful treatment using tacrolimus ointment for cutaneous graft-versus-host disease. *Int J Hematol.* 2008;88(4):465-467. doi: 10.1007/s12185-008-0197-x
56. Dignan FL, Clark A, Amrolia P, Cornish J, Jackson G, Mahendra P, et al. Diagnosis and management of acute graft-versus-host disease. *Br J Haematol.* 2012;158(1):30-45. doi: 10.1111/j.13652141.2012.09129.x.
57. Knobler R, Berlin G, Calzavara-Pinton P, Greinix H, Jaksch P, Laroche L, et al. Guidelines on the use of extracorporeal photopheresis. *J Eur Acad Dermatol Venereol.* 2014;28(Suppl 1):1-37. doi: 10.1111/jdv.12311.
58. Hattori K, Doki N, Kurosawa S, Hino Y, Yamamoto K, Sakaguchi M, et al. Mycophenolate mofetil is effective only for involved skin in the treatment for steroid-refractory acute graft-versus-host disease after allogeneic hematopoietic stem cell transplantation. *Ann Hematol.* 2017;96(2):319-321. doi: 10.1007/s00277-016-2854-0.
59. Yang J, Cheuk DK, Ha SY, Chiang AK, Lee TL, Ho MH, et al. Infliximab for steroid refractory or dependent gastrointestinal acute graft-versus-host disease in children after allogeneic hematopoietic stem cell transplantation. *Pediatr Transplant.* 2012;16(7):771-778. doi: 10.1111/j.1399-3046.2012.01756.x.
60. Tao T, Ma X, Yang J, Zou JY, Ji SM, Tan YS, et al. Humanized anti-CD-25 monoclonal antibody treatment of steroid-refractory acute graft-versus-host

disease: a Chinese single-center experience in a group of 64 patients. *Blood Cancer J.* 2015;5(4):e308. doi: 10.1038/bcj.2015.33.

61. MacMillan ML, Weisdorf DJ, Davies SM, DeFor TE, Burns LJ, Ramsay NK, et al. Early antithymocyte

globulin therapy improves survival in patients with steroid-resistant acute graft-versus-host disease. *Biol Blood Marrow Transplant.* 2002;8(1):40–46. doi: 10.1053/bbmt.2002.v8.pm11858189.M